UNCLASSIFIED

AD NUMBER AD468160 **NEW LIMITATION CHANGE** TO Approved for public release, distribution unlimited **FROM** Distribution authorized to U.S. Gov't. agencies and their contractors; Administrative/Operational Use; Jul 1965. Other requests shall be referred to U.S. Army Biological Labs., Fort Detrick, MD. **AUTHORITY** BDRL ltr, 28 Sep 1971

SECURITY MARKING

The classified or limited status of this report applies to each page, unless otherwise marked.

Separate page printents MUST be marked accordingly.

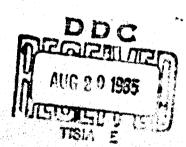
THIS DOCUMENT CONTAINS INFORMATION AFFECTING THE NATIONAL DEFENSE OF THE UNITED STATES WITHIN THE MEANING OF THE ESPIONAGE LAWS, TITLE 18, U.S.C., SECTIONS 793 AND 794. THE TRANSMISSION OR THE REVELATION OF ITS CONTENTS IN ANY MANNER TO AN UNAUTHORIZED PERSON IS PROHIBITED BY LAW.

NOTICE: When government or other drawings, specifications or other data are used for any purpose other than in connection with a definitely related government procurement operation, the U. S. Government thereby incurs no responsibility, nor any obligation whatsoever; and the fact that the Government may have formulated, furnished, or in any way supplied the said drawings, specifications, or other data is not to be regarded by implication or otherwise as in any manner licensing the holder or any other person or corporation, or conveying any rights or permission to manufacture, use or sell any patented invention that may in any way be related thereto.



AD

TECHNICAL MANUSCRIPT 230 TO NATURAL RESISTANCE OF HAMSTER CELLS TO RAZAGUANINE **TO 8-AZAGUANINE**



468160

TECHNICAL MANUSCRIPT 230

NATURAL RESISTANCE OF HAMSTER CELLS TO 8-AZAGUANINE

Alan Richter

JULY 1965



UNITED STATES ARMY BIOLOGICAL LABORATORIES FORT DETRICK Reproduction of this publication in whole or part is prohibited except with permission of Commanding Officer, U.S. Army Biological Laboratories, ATTM: Technical Releases Branch, Technical Information Division, Fort Detrick, Frederick, Maryland, 21701. However, DDC is authorized to reproduce the publication for United States Government purposes.

DDC AVAILABILITY BOTICES

Qualified requestors may obtain copies of this publication from DDC.

Foreign announcement and dissemination of this publication by DDC is not authorized.

Release or announcement to the public is not authorized.

DISPOSITION INSTRUCTIONS

Destroy this publication when it is no longer needed. Do not return it to the originator.

The findings in this publication are not to be construed as an official Department of the Army position, unless so designated by other authorised documents.

U.S. ARMY BIOLOGICAL LABORATORIES Fort Detrick, Frederick, Maryland

TECHNICAL MANUSCRIPT 230

NATURAL RESISTANCE OF HAMSTER CELLS TO 8-AZAGUANINE

Alan Richter

Virus and Rickettsia Division
DIRECTORATE OF BIOLOGICAL RESEARCH

Project 1L013001A91A

July 1965

ABSTRACT

In studies designed to introduce mutations into selected animal cell lines in culture, cell lines derived from the Syrian hamster were found to be naturally resistant to 8-azaguanine. Resistance to the drug was measured by plating colony-forming units at different drug concentrations and determining the concentration at which 50% of the colony-forming units formed colonies. By thi technique it was shown that KB cells (human) and L cells (mouse) have approximately the same susceptibility to 8-azaguanine as reported by others for L, D98 (human), and P388 (mouse). On the other hand, both P113, a polyoma-transformed derivative of embryonic hamster kidney, and BHK21013, an untransformed houseer ceil displaying contact inhibition, proved 50 times more resistant than the cell lines of human and murine origin. The higher resistance of the hamster cells to 8azaguanine is specific, since these cells are still susceptible to the unrelated drug 5-bromodeoxyuridine. Preliminary results suggest that the basis for_natural resistance is not loss of inosinic-guanylic acid pyrophosphorylase activity, because the cells form colonies under conditions in which growth depends on an exogenous supply of hypoxanthine (HAT medium). The naturally resistant hamster cells therefore resemble the mutant D98/AG described by Szybalski.

NATURAL RESISTANCE OF HAMSTER CELLS TO 8-AZAGUANINE

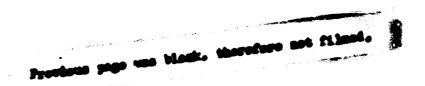
In our laboratory we have been characterizing several animal cell lines for drug susceptibility preparatory to the selection of specific enzyme-deficient mutants. We have been primarily interested in mutants deficient in inosinic-guanylic acid pyrophosphorylase activity that display resistance to 8-azaguanine as a consequence of enzyme loss, and in mutants deficient in thymidine kinese activity that display resistance to 5-bromodeoxymridine.

Susceptibility was determined by plating cells in Eagle's medium containing the drug and counting the number of colonies that formed in the presence and absence of the drug. The number of cells forming colonies in the presence of the drug divided by the number of cells forming colonies in the absence of the drug is expressed in per cent as the relative plating efficiency. By plotting graphically the relative plating efficiency at different concentrations of the drug, it is possible to determine the concentration of the drug that gives a relative plating efficiency of 50% or the LD₅₀ concentration of the drug. Data for the susceptibility of cell lines to 8-azaguanine were available in the literature for mouse strain L cells, mouse strain P388, and human strain DSo. All of these cells were inhibited 50% by drug concentrations between 2 x 10⁻⁷ and 4 x 10⁻⁷ M 8-azaguanine.

We first tested L cells and KB (human) cells. The data are shown in Tables 1 and 2. Each value represents the average of five dishes. By plotting the data of Tables 1 and 2 graphically, it can be shown that the LD₅₀ concentration of 8-azaguanine for L cells is about 2×10^{-7} M and that the LD₅₀ concentration for KB cells is about 4×10^{-7} M. Table 3 compares the data for L cells with those published in the literature for P388 and D98. The LD₅₀ concentrations are similar.

We next examined a hamster cell called P113, which was isolated by Dr. Defendi* after polyoma infection of embryonic hamster kidney. This cell line was unaffected by concentrations of 8-azaguanine that completely eliminated populations of L or KB cells. By employing increasing concentrations of the drug we found that 2 x 10° M 8-azaguanine was necessary to halve the plating efficiency of this cell. To test a possible association between 8-azaguanine resistance and polyoma transformation, we examined a so-called normal hamster cell, BHK21C13. However, the latter proved to be resistant at the same level of drug as the transformed line. This is shown in the fourth table. The increased resistance of the hamster cells to 8-azaguanine is specific, since these cells are still highly susceptible to the unrelated drug 5-bromodeoxyuridine. Preliminary

^{*} Dr. Vittorio Defendi, Wistar Institute, Philadelphia, Pa.



results suggest that the basis for natural resistance is not the loss of inosinic-guanylic acid pyrophosphorylase activity because the cells form colonies under conditions in which growth depends upon an exogenous supply of hypoxanthine in the medium (hypoxanthine, aminopterin, thymidine). Growth in HAT medium is considered to be an indirect test for the enzyme function. It thus seems that the naturally resistant hamster cells resemble the mutant described by Szybalski as D98/AG, which is resistant to 8-azaguanine but possesses the pyrophosphorylase function. Attempts to select enzyme-deficient mutants from the hamster cells are in progress.

7

TABLE 1. EPFECT OF 8-AZAGUANINE ON THE RELATIVE PLATING EFFICIENCY OF L CELLS

	Cell Ir	Cell Input, 90 per dish	itsh	Cell In	Cell Input, 180 per dish	di sh
M Concentration of Drug	Average No. Colonies Found Per Dish	Plating Efficiency	Relative Plating Efficiency	Average No. Colonies Found Per Dish	Plating Efficiency	Relative Placing Efficiency
0	51.8	57.5	100	88	48 .8	100
1 × 10'	48.6		76	72.8		82.7
3 x 10°	17.4		33.6	24.4		27.7
5 x 10"	0.2		0.39	0.2		0.23
8 x 10.	0			0		
1 × 10.	0			0		

TABLE 2. EFFECT OF 8-AZAGUANINE ON THE RELATIVE PLATING EFFICIENC: OF KB12 CELLS

	Cell In	Cell Input, 90 per dish	sh	Cell Inc	Cell Input, 180 per dish	dish
M Concentration of Drug	Average No. Colonies Found Per Dish	Flating Efficiency	Relative Plating Efficiency	Average No. Colonies Found Per Dish	Plating Efficiency	Relative Plating Efficiency
0	50.7	61.1	100	119.5	62.2	100
1 x 10 ⁻⁷	61.4		104	93.0		78.0
3 x 10"	42.0		71.6	58.8		49.2
5 x 10°	30.4		51.9	5.2		4.4
8 x 10"	4.6		16.0	0		(<0.8%)
1 × 10.	4.2		7.2	0		(<0.8%)

TABLE 3. SUSCEPTIBILITY OF DIFFERENT CELL LINES TO 8-AZAGUANINE

Deug	Relative Plating Efficiency, % survival			
Drug Concentration	L	P388ª/	D98 <u>b</u> /	
1 x 10 ⁻⁷ M	88.4	90	•	
3 x 10 ⁻⁷	30.7	59	-	
5 x 10 ⁻⁷	0.31	10	-	
8 x 10 ⁻⁷	0	0	50	
1 x 10 ⁻¹⁰	0	0	•	

a. Information from Bradley et al.1

TABLE 4. DRUG SUSCEPTIBILITY OF CELL STRAINS

	LD ₈₀ Concentration of Drug ^a /			
Cell	8-azaguanine, 10 ⁷⁷ molar	5-bromodeoxyuridine, 10 molar		
Mouse strain L	4.5	17.0		
Human strain KB	4	8.5		
Syrian hamster strain 113	200	2.5		
внк21С13	200	ND <u>b</u> /		

a. Concentration of drug permitting 50% relative plating efficiency compared with plating in absence of drug. All determinations in Eagle's medium containing 10% calf serum.

b. Information from Szybalska and Szybalski.

b. ND - Not determined.

LITERATURE CITED

- 1. Bradley, T.R.; Roosa, R.A.; Law, L.W. 1962. DNA transformation studies with mammalian cells in culture. J. Cellular Comp. Physiol. 60:127-137.
- 2. Szybalska, E.H., Szybalski, W. 1962. Genetics of human cell lines: IV. DNA-mediated heritable transformation of a biochemical trait. Proc. Nat. Acad. Sci. 48:2026-2034.
- 3. Macpherson, I., Stoker, M. 1962. Polyoma transformation of hamster cell clones an investigation of genetic factors affecting cell competence. Virology 16:147-151.

DOCUMENT CO	NTROL DATA - R&	D	he averall report is a lassified.
(Security classification of title, Judy of abstract and index	ING MITTHEFFE THE WATER OF	2 M HE POF	PT SECONITY CLASSIFICATION
		Une l	lassified
v.s. Army Biological Laboratories Fort Detrick, Frederick, Maryland, 21	701	Zh GROUP	
The period of th			
3 REPORT TITLE			
NATURAL RESISTANCE OF HAMSTER CELLS TO	O 8-AZAGUANINE		
4 DESCRIPTIVE NOTES (Type of report and inclusive dates)			
5 AUTHORISS (Last name, first name, initial)			. T
Richter, Alan			
6 REPORT DATE	78 TOTAL NO OF P	AGES	75 NO. OF REFS
July 1965	14		3
SH CONTRACT OR GRANT NO.	9# ORIGINATOR'S R	EPORT NUM	BER(5)
h PROJECT NO 11.013001A91A	Technical M	anuscrip	ot 230
c	9b. OTHER REPORT	NO(S) (Any	other numbers that may be sasigned
d Notice	<u> </u>		
Qualified requestors may obtain copies Foreign announcement and dissemination Release or announcement to the public	of this public	ation by	
11 SUPPLEMENTARY NOTES	12. SPONSORING MILI	TARY ACTI	VITY
		_	l Laboratories Prick, Maryland, 21701
13 ASSTRACT			

In studies designed to introduce mutations into selected animal cell lines in culture, cell lines derived from the Syrian hamster were found to be naturally resistant to 8-azaguanine. Resistance to the drug was measured by plating colonyforming units at different drug concentrations and determining the concentration at which 50% of the colony-forming units formed colonies. By this technique it was shown that KB cells (human) and L cells (mouse) have approximately the same susceptibility to 8-azaguanine as reported by others for L, D98 (human), and P388 (mouse). On the other hand, both P113, a polyoma-transformed derivative of embryonic hamster kidney, and BHK21C13, an untransformed hamster cell displaying contact inhibition, proved 50 times more resistant than the cell lines of human and murine origin. The higher resistance of the hamster cells to 8azaguanine is specific, since these cells are still susceptible to the unrelated drug 5-bromodeoxyuridine. Preliminary results suggest that the basis for natural resistance is not loss of inosinic-guanylic acid pyrophosphorylase activity, because the cells form colonies under conditions in which growth depends on an exogenous supply of hypoxanthine (HAT medium). The naturally resistant hamster cells therefore resemble the mutant D98/AG described by Szebalski.